

Product no **AS08 300****1 | PEB (4x) | protein extraction buffer****Product information**

Quantity	5 x 2 ml (4x stock) allows up to 75 isolations of plant material (using 500 µl 1x PEB for 100 mg fresh weight) or 190 isolations of algal material (using 200 µl 1x PEB for cell amounts corresponding to 4-10 µg total chlorophyll)
Storage	Stable at RT for at least 1 month; short-term storage (6 monthss) at 4 °C and long term storage (1 year or more) at -20 °C.
Additional information	<p>Buffer components (4x): contains ~ 40% v/v glycerol [HOCH₂CH(OH)CH₂OH], Tris-HCl [NH₂C(CH₂OH)₃ · HCl] pH 8.5, LDS [CH₃(CH₂)₁₁OSO₃Li], EDTA [(HO₂CCH₂)₂NCH₂CH₂N(CH₂CO₂H)₂]</p> <p>It is recommended to include a protease inhibitor (not supplied with this buffer) from a freshly made stock while preparing the ready-to-use 1x PSB.</p> <p>PEB has been optimized for quantitative small-scale preparation of whole protein extracts from plant/algal tissue. Extraction using the procedure described below will result in maximum yield of proteins and diminish protein degradation and aggregation.</p> <p>Extracts may be quantified using detergent (LDS) compatible methods and have been shown to give highly reproducible and quantitative results in subsequent SDS PAGE gel electrophoresis, Western Blotting, and immunoprecipitation.</p> <p>PEB has been tested on a wide range of species and tissues from higher plants, mosses, lichens, algae, diatoms, dinoflagellates, and cyanobacteria.</p>

Application information

Confirmed reactivity	<p>PEB has been tested on a wide range of species and tissues from Higher plants, Mosses, Lichens, Algae, Diatoms, Dinoflagellates, Cyanobacteria. Extracts may be quantified using detergent (LDS) compatible methods, and have been shown to give highly reproducible and quantitative results in subsequent SDS PAGE gel electrophoresis, Western blotting, and Immunoprecipitation (IP).</p> <p>Most of Agrisera commercial antibodies are tested on plant or algal samples extracted with this buffer. An example can be found here.</p>
Selected references	<p>Altuntas et al. (2020). Proline-stimulated signaling primarily targets the chlorophyll degradation pathway and photosynthesis associated processes to cope with short-term water deficit in maize. <i>Photosynth Res.</i> 2020 Apr;144(1):35-48. doi: 10.1007/s11120-020-00727-w.</p> <p>Pérez-López et al. (2020). Transcriptome Analysis Identifies Plasmodiophora brassicae Secondary Infection Effector Candidates. <i>J Eukaryot Microbiol.</i> 2020 Jan 11. doi: 10.1111/jeu.12784.</p> <p>Morin et al. (2019). Morin et al. (2019). Response of the sea-ice diatom <i>Fragilariopsis cylindrus</i> to simulated polar night darkness and return to light. <i>Limnology and Oceanography.</i> 9999, 2019, 1-20. (sea-ice diatom)</p> <p>Bausch, A.R., Juhl, A.R., Donaher, N.A. et al. (2019) 166: 80.</p> <p>Matsuo and Atsumi (2018). Xylosylation of proteins by expression of human xylosyltransferase 2 in plants. <i>J Biosci Bioeng.</i> 2018 Sep;126(3):371-378. doi: 10.1016/j.jbiosc.2018.03.013.</p> <p>Brouwer et al. (2011) TheImpact ofLightIntensity onShade-InducedLeaf Senescence. <i>Plant Cell Environ.</i> Dec. 15 (ahead of print).</p> <p>Kosawang et al. (2011) Hydrogen yield from a hydrogenase in Frankia R43 at different levels of the carbon source propionate. <i>Journal of Environmental Management,</i> Jan 26</p>

Before you start

Prepare sufficient 1x PEB for all samples by diluting 4x stock in sterile, deionized water (the pH of your 1x PEB should be between 8.25 and 8.75). It is recommended to include a protease inhibitor (not supplied with this buffer) from a freshly made stock while preparing the ready-to-use 1x PEB to increase the yield of non-degraded protein in the extract. We recommend including 1:50 vol/vol from a freshly prepared 50x stock (in 1x PEB) to give the desired final concentration recommended by the manufacturer (e.g. 0.1 mg/ml for Pefabloc SC, Roche).

The total volume of 1x PEB required is dependent on the sample type and amount of tissue used: for 100 mg fresh plant tissue we recommend 500 µl 1x PEB; for algal samples (corresponding to 4-10 µg total chlorophyll) we recommend 200 µl 1x PEB. Keeping sample volumes in a range of 0.2-0.5 ml has been found to contribute to better extraction results, an upscale in volume is not recommended.

Material preparation

Plant tissue: weigh and snap freeze in liquid nitrogen and store at -80°C until processing.

Algal cultures: centrifuge to form a pellet or collect on filters (e.g. GF/F or polycarbonate) and freeze at -80°C until processing.

Extraction

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| 1 | Grind frozen material in liquid N ₂ in a pre-chilled mortar with a pestle to a fine powder and transfer to a 1.5 ml tube | Keep material cool at any time during grinding, avoid spillage |
| 2 | Add 1x PEB and immediately freeze sample in liquid N ₂ | 500 µl for 100 mg plant tissue or 200 µl for cells corresponding to 4-10 µg total chlorophyll; keep tube upright to hold sample at the bottom of the tube |
| 3 | Carefully subject sample to sonication just until sample is thawed, re-freeze sample immediately in liquid N ₂ to avoid heating | Optimal results will be obtained using a microtip sonicator (e.g. Branson Ultrasonics Model 450) at low settings of about 30%; waterbath sonicators may also be used though this may lead to slightly less reproducible protein recovery rates; |
| 4 | Repeat sonication step (3) depending on species, place on ice until all samples are processed | For higher plants 2-3 cycles, for cyanobacteria 3 cycles, for <i>Chlamydomonas</i> 2 cycles, for <i>Heterosigma</i> , <i>Thalassiosira</i> and <i>Trichodesmium</i> 1 cycle |
| 5 | Centrifuge your samples for 3 min at 10 000 x g to remove insoluble material and unbroken cells, the pellet should be white/light-grey | An intense green color of the pellet indicates that disruption was not optimal and extraction conditions need to be adjusted (e.g. improved grinding and/or repeated sonication steps) using a new sample |
| 6 | Transfer supernatant to new tube using a pipette, be careful not to carry over debris | Expect ~400 µl supernatant for the plant and ~150 µl for cyanobacterial/algal samples; collecting supernatant with a pipette as 2 x 200 (or 2 x 75 µl) reduces the risk of disturbing the pelleted debris |

Protein determination

Assay total protein content of recovered supernatant using a detergent compatible assay. Based on the amount and/or tissue of the species used you may expect a protein content of 1.5-6 µg/µl.

Storage

Protein extracts may be stored for 24 hrs at +4°C or up to 6/12 month at -20°C/-80°C. We recommend to aliquot samples. Re-freezing protein samples may induce degradation/aggregation.

Loading on a gel

A freshly prepared reducing agent should be added (e.g. Dithiothreitol, final concentration 50 mM) to the volume prepared for loading. Heat at 70°C for 5 min, briefly spin down and load on a gel. Protein loads of 0.5-5 µg/lane should be sufficient for most Western Blot applications.